1	The Role of Prothrombin Gene and Methylenetetrahydrofolate Reductase
2	Gene Polymorphisms and Thrombophilia Markers as Risk Factors for
3	<b>Recurrent unexplained Miscarriage</b>
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## Abstract

Background: Recurrent unexplained miscarriage is still an unsolved reproductive health 15 problem. Inherited thrombophilias have been accused as one of the causes. Secondary to an 16 increased tendency for venous thromboembolism because of a mutation in a gene encoding a 17 protein involved in the coagulation cascade. These include prothrombin gene (PT G20210A) and 18 methylenetetrahydrofolate reductase (MTHFR) mutations. The study aims to evaluate the 19 association between polymorphisms in the prothrombin gene and the MTHFR gene with 20 recurrent miscarriage (RM). We also evaluated the association between Protein C (PC). Protein S 21 (PS), Antithrombin III (ATIII), and homocystiene with recurrent miscarriage (RM). 22

Methods: We conducted a comparative study on women with a history of two or more miscarriages and healthy controls with no history of miscarriage and who had at least completed one full-term normal pregnancy. Genetic analysis of the participants was done using the 5' Nuclease Assay (TaqMan) PCR technique and various other blood tests were performed to check general health indicators and thrombophilia markers.

Results: In this study of 195 RM group (Group I) participants and 90 healthy controls (Group 28 II), we noted significant discrepancies in health conditions. PC deficiency occurred in 7.2% of 29 Group I, but only 1.1% of Group II. PS deficiency was found in 65.6% of Group I versus 7.8% 30 of Group II. ATIII deficiency was observed in 9.2% of Group I and 2.2% of Group II. 31 Hyperhomocysteinemia was noted in 10.8% of Group I, and 2.2% of Group II. For the 32 prothrombin gene G20210A, two Group I participants were A/G, with no A/G in Group II, and 33 no AA carriers in either group. G allele was in 99.5% of Group I and 100% of Group II, while 34 the A allele was in 0.5% of Group I only. MTHFR C677T gene showed C/T mutation in 33.3% 35 36 of Group I and 32.2% of Group II, and T/T mutation in 12.8% of Group I and 8.9% of Group II. The C allele was found in 70.5% of Group I and 75% of Group II, with the T allele in 29.5% of 37 38 Group I and 25% of Group II (p=0.269).

**Conclusion**: Prothrombin gene G20210A and MTHFR C677T gene polymorphisms are not correlated with RM in the Egyptian population. About 70% of women in upper Egypt have at least one type of MTHFRC677T gene polymorphism. However, Egyptian women with RM are strongly associated with hyperhomocysteinemia, PC, PS, and AT deficiencies.

- 43 Keywords: Prothrombin gene; Methyltetrahydrofolate reductase; Thrombophilia markers;
- 44 Recurrent miscarriage.

## 46 **1. Background**

The exchange of nutrients, gases, and other metabolites during pregnancy depends on the 47 connection between the placenta and the maternal circulatory system. Recurrent miscarriages 48 (RM) may be caused by abnormal blood coagulation in the small blood vessels of the placenta 49 (1). American Society for Reproductive Medicine (ASRM) defines RM as "two or more failed 50 clinical pregnancies" (2). Inherited thrombophilia is characterized by an increased tendency for 51 venous thromboembolism as a result of a mutation in a gene encoding a protein involved in the 52 coagulation cascade. These include methylenetetrahydrofolate reductase (MTHFR) mutation, 53 antithrombin III (ATIII) deficiency, protein C and protein S deficiency (PSD), prothrombin gene 54 (PT G20210A) mutation, and factor V Leiden (FVL) (3). In the prothrombin G20210A variant, 55 adenine replaces guanine at nucleotide position 20210 in the 3' untranslated region of the gene. 56 Increases in the amount and activity of prothrombin in blood plasma are associated with the GA 57 genotype and increased risk of thromboses (4). The MTHFR gene encodes MTHFR, the rate-58 limiting enzyme in the methyl cycle. C677T (rs1801133) and A1298C (rs1801131) are two of 59 the most extensively studied single nucleotide polymorphisms (SNPs) (5). 60

Deficient MTHFR enzyme activity, often the result of inherited mutations, is a leading 61 62 cause of hyperhomocysteinemia (6). In patients with a previous abortion history, MTHFR polymorphism was shown to have a substantial effect. Hyperhomocysteinemia, leading to a 63 hypercoagulable condition, is the leading cause of early pregnancy loss. Sperm quality and 64 quantity, as well as reduced ovarian reserve, are affected by MTHFR isoforms (7). Liver cells 65 produce a glycoprotein called protein C (PC) that requires vitamin K for proper functioning. 66 Disseminated intravascular coagulation or massive thrombosis are the two main manifestations 67 of a PC deficiency (8). A seven-fold increase in thrombotic risk was observed in patients with 68

PC deficiency. While in terms of protein S (PS), a vitamin K-dependent glycoprotein, the risk 69 ratio of thrombosis is 8.5 times more than in normal individuals (9). Women with PS deficiency 70 are more likely to have a VTE during pregnancy or puerperium, and their risk of RM is three 71 times higher than that of the general population (10). The vitamin K-independent glycoprotein 72 antithrombin (AT) is essential in the clotting cascade (11). Approximately 60% of cases with AT 73 74 deficiency occur spontaneously, and this condition is passed down in an autosomal dominant manner, making carriers more likely to have thrombosis and VTE. If a woman has a history of 75 VTE, her risk of developing a thrombus during pregnancy rises from 31% to 50% because of her 76 77 AT deficit (12). The current study aims to evaluate the association between polymorphisms in the prothrombin gene and the MTHFR gene with RM among the Egyptian population. 78 Additionally, we studied the prevalence of thrombophilia markers, including Homocysteine, PC, 79 PS, and AT III, in patients with RM in Upper Egypt. 80

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## 2. Materials and Methods

## 82 Study Design and Patients:

A comparative study pre-registered study (NCT03209063) recruited women from the 83 Clinical Pathology Department, Woman's Health Hospital, Assuit University Hospital, Assuit 84 University during the period from December 2019 through May 2022. Assuit Medical School, 85 Ethical Review Board, had approved the study number (17200095). We recruited women in the 86 87 intervention group who were diagnosed to have RM (as defined by the ASRM) as "two or more failed clinical pregnancies" and accepted to participate in the study and were less than 35 years 88 old. Conversely, the control group consisted of healthy females under 35 who had no miscarriage 89 90 history and at least one problem-free full-term pregnancy. Women with unregulated diabetes

91 mellitus, hyperthyroidism, autoimmune conditions like antiphospholipid antibody syndrome with
92 a LA1/LA2 ratio exceeding 1.2, and those presently using oral contraceptives or anticoagulant
93 treatments were excluded from both study groups.

The study protocol had been approved by Assiut University Ethical Review Board. We affirmed that all study procedure complies with the Declaration of Helsinki principles. All women signed the written informed consent before enrollment. We followed the STROBE guidelines while drafting this manuscript (13).

# 98 Data Collected and Sampling:

Eligible participants had an interview with the investigator during which the following
data were collected included age, menstrual history, obstetric history (prenatal, natal, postnatal),
number of previous abortions, gynecological history, contraceptive history, family history
(history of abortion in the family, history of DVT), history of operations and drug history were
specifically considered.

From each participant and control subject, approximately ten ml of venous blood was 104 collected in a fully sterile environment. The collected sample was then distributed as follows: 105 one ml was placed in an EDTA-containing tube for a complete blood count (CBC), two ml in 106 another EDTA-containing tube for genotyping, and three ml in an anticoagulant-free tube for 107 random blood glucose, kidney and liver function tests, and thyroid stimulating hormone (TSH) 108 109 evaluation. The remaining 4 ml of blood was put into citrated tubes for the measurement of PC, PS, and ATIII. The CBC and kidney/ liver function tests were performed using the CELL-DYN 110 RUBY (Abbott - USA) and Cobas c311 (Roche - Germany). The coagulation profile was 111 assessed using the auto analyzer Sysmex CA- 1500 (Siemens -Germany), while the serum TSH 112

113 was analyzed using Vidas (Biomerieux-France). The PC, PS, and ATIII were analyzed by auto-

114 analyzer Sysmex CA- 1500 (Siemens –Germany). Homocystiene was analyzed by ADVIA

115 Centaur XPT (Siemens -Germany).

In the genetic analysis, DNA was isolated from plasma samples for the evaluation of
Prothrombin gene G20210A and MTHFR C677T gene single nucleotide polymorphisms (SNP).
The extraction was carried out using a real-time PCR Fast 7500 Applied Biosystems and a
Genejet Whole Blood genomic DNA purification mini kit -Cat. No. (K0781) provided by
Thermo Fisher Scientific, Waltham, MA, USA. To ensure consistency, all DNA samples were
diluted with nuclease-free water to yield a concentration between 1-20 ng per well, with a
recommended minimum final concentration of 0.2 ng/μL.

The study employed the 5' Nuclease Assay (TaqMan) PCR technique. This method 123 involves a specific TaqMan MGB probe annealing to its complementary sequence between the 124 forward and reverse primer sites. The AmpliTag Gold DNA polymerase, with its 5' nuclease 125 activity, cleaves probes that have hybridized to the target sequence, separating the quencher dye 126 from the reporter dye and leading to an increase in fluorescence. This fluorescence indicates 127 which alleles are present in the sample. The PCR process involves preparing the reaction mix, 128 129 DNA samples, and the reaction plate, followed by running the PCR and post-PCR analysis on a real-time PCR instrument. The software of the real-time PCR instrument analyzed fluorescence 130 131 signals from each well, which are represented as Rn values, to identify the specific alleles in each 132 sample. These results are obtained from the amplification reactions performed during the Allelic 133 discrimination genotyping assay.

134 Study outcomes:

The primary outcome of this study was to examine the occurrence of prothrombin gene and MTHFR gene polymorphisms in women with recurrent miscarriages in comparison to a healthy control group. In addition, secondary objectives included detecting thrombophilia markers such as PC (normal range: 70-140%), PS (normal range: 60-130%), ATIII (normal range: 79.4-112%), and Homocysteine (normal range: 3.7 to 13.9 µmol/L) in both groups of women..

## 140 Sample size and data analysis:

Taking the percentage of patients with RM with prothrombin gene polymorphism as a 141 primary outcome. Previous studies showed that the percentage was (10.9%) (Salim Sehirali et 142 al., 2005). Additionally, the percentage of this genetic polymorphism in the control healthy 143 population was (1.06%) (Arzu Ulu et al., 2006). Considering a confidence level of 95%, 80% 144 power, and recruited 180 patients, 90 in each group ratio of 1:1, with a risk ratio of 0.09 (Epinfo 145 2019). However, due to the scarcity of positive cases in the interim of data analysis, we doubled 146 the sample size of cases to perform our analysis in 195 cases and 90 controls. Data were 147 analyzed using the Statistical Package for Social Science (SPSS) version 25 software for 148 Windows. Statistics were generated for categorical data in the form of frequencies and 149 percentages, and Shapiro–Wilk tests were used to assess the normality of numerical variables, 150 151 presenting these as either median (range) for non-normal distribution or mean  $\pm$  SD for normal distribution. Comparative analyses between groups were conducted using Chi-square, Fisher 152 153 Exact tests, and Mann Whitney U tests for proportions and median differences, while Spearman's 154 correlation was utilized to find relationships between continuous variables. A P value less than 155 0.05 was deemed statistically significant.

156 **3. Results** 

We recruited 195 cases in the RM group (Group I) and 90 controls (Group II) (Figure 1). 157 The median age was 26 (16-45) years among Group I and 27 (19-35) years among Group II. 158 Nearly 52.3% and 52.2% were urban in Group I and Group II, respectively. Besides, 9.2% of 159 Group I and 4.4% of Group II have a family history of VTE. No statistically significant 160 161 difference between Group I and Group II in all previous parameters. About 44% have a history of > 2 previous abortions. Regarding the time of previous abortion, in the first abortion, 75.9% 162 occurred in  $\leq 12$  weeks. In second abortion, 79.9% occurred in  $\leq 12$  weeks. Of more than three 163 abortions, 77.8% occurred in  $\leq 12$  weeks and 22.2 % in > 12 weeks (**Table 1**). 164

In terms of laboratory characteristics, the median range of WBC count in Group I was 5.9 165 (2.47-13.60), compared to 5.79 (2.47-10.50) in Group II. The median prothrombin time level in 166 Group I was 12.20 (10.1-15.4), and in Group II was also 12.20 (10.5-14.5). The median APTT in 167 Group I was 31.6 (22.5-43.1) second, and in Group II was also 31.65 and ranged from 22.5-43.1. 168 169 The median TT in Group I was 18.00 (12.2-35.0), and in Group II was also 17.85 (12.2-35.0). Lastly, the median Fibrinogen in Group I was 3.20 (1.7-6.2), and in Group II was also 3.20 (2.0-170 5.8). No statistically significant difference between Group I and Group II in all previous 171 172 parameters, Table 2.

PC deficiency in Group I was 7.2% compared to 1.1% in Group II.PS deficiency in Group I was 65.6% compared to 7.8% in Group II. ATIII deficiency in Group I was 9.2% compared to 2.2% in Group II. Hyperhomocysteinemia in Group I was 10.8% compared to 2.2% in Group II. There was a statistically significant difference between Group I and Group II in all the above parameters (**Figure 2**).

In terms of prothrombin gene G20210A, the wild G/G presented in 99.0 % of Group I 178 and in 100.0 % of Group II, while Hetero-mutant A/G presented in only two patients among 179 Group I compared to no participants among Group II. There was no AA either in Group I or 180 Group II. Allel G presented in 99.5% of Group I and 100.0 % of Group II, while Allel A 181 presented in 0.5% of Group I and not presented in Group II. Regarding the MTHFR C677T gene, 182 wild C/C presented in 53.8% of Group I and in 58.9 % of Group II, Hetero-mutant C/T presented 183 in 33.3% of Group I and in 32.2% of Group II, and Homomutant T/T presented in 12.8% among 184 Group I and in 8.9% among Group II. Allel C presented in 70.5% of Group I and in 75 % of 185 Group II, while Allel T presented in 29.5% of Group I and in 25% of Group II. No statistically 186 significant difference in the distribution of Prothrombin gene G20210A, MTHFR C677T gene 187 and their alleles between Group I and Group II, (Table 3). 188

## 189 **4. Discussion**

190 In this comparative study, we could not find an association between prothrombin gene 191 G20210A or MTHFR C677T gene polymorphisms and RM in the population of Upper Egypt. On the other hand, thrombophilia markers were positively correlated with RM as compared to 192 193 the healthy population. Although age may affect the incidence of RM; however, our eligibility criteria were limited to less than 35 years, which nullifies the effect of age on RM. The rate of 194 family history of VTE ranged from 4.4% and 9.2%, with no statistically significant difference 195 between cases and control groups. In the presented study, about 56% of the cases had a history of 196 two previous abortions, and 44% had a history of >2 previous abortions. Research conducted by 197 Ogasawara and his team noted a rise in the rate of miscarriages from 25% to 80% as the number 198 of previous miscarriages increased from 2 to 7 or more, predominantly due to a growing 199

200 occurrence of miscarriages with a standard karyotype. On the other hand, the incidence of 201 miscarriages with chromosomal abnormalities remained consistent (14).

202 The prevalence of PC deficiency in our RM patients was substantially greater than in the 203 control group (7.4% vs 1.1%, respectively). Likewise, Jyotsna et al. showed a statistically significant increase in PC deficit among RM patients compared to healthy controls (33.3% vs 204 205 3.3%, respectively) (15). Hansda and Roychowdhury conducted another Indian investigation on 53 RM cases and 47 healthy age-matched controls, and they found that 15.09% of the RM 206 patients had a deficit PC (16). On the other hand, Osman and Abulata could not detect a 207 statistically significant difference in PC levels in RM patients and control (8). Our results differ 208 from those of the Osman and Abulata study, but this may be because we recruited a much larger 209 sample size and included women who experienced both first- and second-trimester RM, whereas 210 they only included women who had experienced RM in the first trimester. 211

Sixty-five percent of our recruited patients in the RM group were shown to have a 212 significantly lower level of PS compared to 7.8% in the control group. Similarly, PS deficiency 213 was observed to be more common in RM patients than in controls by both Alshammary et al. and 214 Jyotsna et al. (15.17). Parand et al. found a significant association of RM with PS deficiency in a 215 sample of 90 patients who had experienced three or more consecutive miscarriages with the same 216 partner at less than 20 weeks of gestation (18). A study by Matsukawa et al. on 355 Japanese 217 218 women with RM and 101 parous women indicated that PS deficiency did not act as a reliable clinical predictor of RM (19). The discrepancy between their findings and ours might be 219 attributed to the fact that lupus anticoagulant was not taken into account in their research. In 220 221 addition, Mekaj et al. observed no statistically significant difference between 104 cases of RM in

the first trimester and 110 controls in their research conducted in Kosovo (20). Disagreements 222 with our findings may be attributable to differences in study population age and ethnicity. 223

224 Deficiency in AT was observed in 9.2% of the RM group vs. 2.2% in the control group. 225 This finding is consistent with that of Jyotsna et al., who also reported a statistically significant correlation between the mean value of AT in the patient and control groups (15). On the other 226 227 hand, Mekaj et al. did not find a significant difference in terms of AT deficiency between the RM and the control groups (20). In the present study, we found a statistically significant higher 228 level of Homocysteine in the RM group than in controls, as hyperhomocysteinemia in cases with 229 RM was 10.8% compared to 2.2% in the control group, and it was statistically significant 230 (p=0.017), in harmony with Abd-Ellatef et al. study as the mean homocysteine level was higher 231 in RM group than the control group (21). In the same line, Nelen et al. and Klai et al. found that 232 elevated homocysteine level was a risk factor for recurrent early pregnancy losses (22,23). 233 During pregnancy, levels of homocysteine typically decrease. High concentrations of 234 235 homocysteine might be linked with fetal abnormalities and potential issues with the blood vessels in the placenta, which could lead to abruption (24). There's a growing understanding of 236 the role of increased homocysteine as an independent risk factor for both arterial and venous 237 thrombosis. The suggested pathogenic mechanisms include elevated levels of asymmetric 238 dimethylarginine, impaired methylation, oxidative damage to the endothelium due to suppression 239 of the vasodilator nitric oxide, promoting platelet activation and aggregation, vascular smooth 240 muscle proliferation, and disruption of the usual balance between procoagulants and 241 anticoagulants, favoring thrombosis (25). 242

Regarding prothrombin gene G20210A, we could not find any significant difference 243 between both groups. Similarly, Ashour and Sharif demonstrated that the A allele was more 244 prevalent in RM patients (2.25%) as compared to controls (0.75%) (26). Parand et al. showed 245 that there was no homozygous case for the prothrombin G20210A polymorphism sample (18). 246 Osman and Abulata identified the prothrombin gene G202010A mutation in 11% of case 247 subjects and 2% of control subjects, but the difference was not statistically significant (8). 248 Similarly, Nassour-Mokhtari and colleagues discovered the prothrombin G20210A mutation in 249 both recurrent miscarriage (RM) and control groups, in a heterozygous form, and found no 250 251 association between this mutation and RM (27). The difficulty in identifying a link between this polymorphism and RM may be due to the low frequency of the minor A-allele in our Group. 252 Results may vary if the sample size is larger. Warren et al., who investigated whether women 253 with the G20210A mutation in prothrombin were at increased risk of RM, came to the same 254 conclusion. For this study, investigators enrolled 5188 pregnant women and reviewed the results 255 of 4167 blood samples collected during the first trimester to screen for the G20210A gene 256 mutation. The conclusion of their study revealed no association between the prothrombin gene 257 mutation G20210A and recurrent miscarriage (28). However, a comprehensive meta-analysis 258 259 incorporating 37 case-control studies indicated an elevated risk of recurrent miscarriage linked to the G20210A prothrombin mutation, particularly noticeable in European women and those aged 260 above 29 years (29). 261

Our research showed no significant disparity in the MTHFR C677T gene, including Wild C/C, Hetero-mutant C/T, and Homomutant T/T, between the two groups. Furthermore, there was no significant difference in the frequencies of Allele C and Allele T in both groups. A separate study involving 133 women with a history of three or more consecutive miscarriages before the

20th week of pregnancy revealed allele frequencies for T at 34.6% for cases and 21.6% for the 266 control group, with higher homocysteine levels noted in women carrying the mutant alleles (30). 267 As for genotype frequencies, Fard et al. noted that 30% of the recurrent miscarriage group 268 carried the TT genotype for the MTHFR 677T gene, in contrast to 8% in the control group. 269 Furthermore, 40% of women in the recurrent miscarriage group exhibited a CT genotype, 270 271 associated with elevated homocysteine levels compared to the control group (31). However, a meta-analysis by Chen et al., including 16 articles, found that MTHFR C677T was substantially 272 related to RM risk in the Chinese population across all genetics models (32). In previous studies, 273 the MTHFR C677T polymorphism has been linked to an increased risk of RM. A study by Luo 274 et al. on 136 women with a history of two or more spontaneous abortions found that women with 275 the MTHFR 677T gene may be more likely to experience RM. According to their findings, 276 people with a CT or TT genotype should increase their consumption of folic acid supplements to 277 avoid miscarriage, and MTHFR C677T might serve as an early genetic screening signal for RM 278 (33). It is possible that folic acid supplementation during pregnancy, particularly in the first 279 trimester, had something to do with this. Homozygotes for the MTHFR gene have a much lower 280 homocysteine threshold, and this is in large part due to their elevated folate levels. This led 281 282 researchers to hypothesize that women with two MTHFR mutation variants would benefit from taking folic acid to mitigate the negative consequences of this mutation and lower their risk of 283 284 RM (34). Zhu et al. investigated MTHFR polymorphisms in 370 Chinese women with RM and 285 found that the MTHFR C677T variant was more common in this population (35). In their analysis of 100 Iranian women, Fard et al. found that the MTHFR C677T variant was much more 286 common among those with RM, suggesting that it could be a risk factor for miscarriage (31). In 287 288 addition, Osman and Abulata reported that MTHFR was the most frequently detected gene

deficiency in both the case and control groups (63% and 41.9%, respectively) (8). However, the limited sample size and the use of an unconventional approach (FV-PTH-MTHFR strip assay) might account for these discrepancies in pregnancy outcomes. According to a study by Bigdeli et al., the homozygous frequencies of MTHFR C677T mutations were also elevated among Iranian women who had RM (36). The discrepancy across studies might be due to factors like the diversity of the studied populations, sample sizes, and research designs or methods.

We acknowledge that our study has some limitations, including the lack of adequate data to perform a subgroup analysis regarding prothrombin gene G20210A polymorphism. Further, we could not perform more extensive studies of MTHFR variants and other genetic mutations that may be associated with RM due to the limited financial resources. Additionally, we did not measure vitamin B12 or folic acid serum levels in patients with hyperhomocysteinemia. Multicenter trials with larger sample sizes and different ethnic groups are required.

## 301 Conclusion

Prothrombin gene G20210A and MTHFR C677T gene polymorphisms are not correlated with RM in the Egyptian population. About 70% of women in upper Egypt have at least one type of MTHFRC677T gene polymorphism. However, Egyptian women with RM are strongly associated with hyperhomocysteinemia, PC, PS, and AT deficiencies.

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# 309 **Conflict of interest**: None.

# 310 Authors' contributions

- 311 Z.A.; contributed to protocol development, interpretation of the data, and data analysis. H.G. and
- 312 O.M.S. suggested the research idea, were responsible for the study conception and design and
- revised the manuscript. M.I.S.; and A.A.; contributed to the study design, interpretation of the
- data, and data analysis. T.A.F; and G.M; Participated in data collection, data analysis, and
- manuscript writing. All authors performed editing and approving the final version of the
- 316 manuscript for submission.

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408

Variables	Group I (n=195)	Group II (n=90)	P-Value
Age (years)			
• < 20	10 (5.1%)	2 (2.2%)	0.525*
<b>20-30</b>	118 (60.5%)	56 (62.2%)	_
■ > 30	67 (34.4%)	32 (35.6%)	
Median (range)	26.0 (16-35)	27.0 (19-35)	0.080**
Residence			
<ul> <li>Urban</li> </ul>	102 (52.3%)	47 (52.2%)	0.989*
<ul> <li>Rural</li> </ul>	93 (47.7%)	43 (47.8%)	
Consanguinity			
• Yes	58 (29.7%)	18 (20.0%)	$0.084^{*}$
<ul> <li>No</li> </ul>	137 (70.3%)	72 (80.0%)	-
Family history of VTE			
• Yes	18 (9.2%)	4 (4.4%)	0.159*
<ul> <li>No</li> </ul>	177 (90.8%)	86 (95.6%)	1
Number of previous abortions			
2 abortions	109 (55.9%)		
> 2 abortions	86 (44.1%)		
Time of abortion			
First abortion			
$\leq 12$ weeks	148 (75.9%)		
> 12 weeks	47 (24.1%)		
Second abortion (n=194)			
$\leq 12$ weeks	155 (79.9%)		
> 12 weeks	39 (20.1%)		
Third abortion (n=87)			
$\leq 12$ weeks	75 (86.2%)		
> 12 weeks	12 (13.8%)		
More than 3 (n=36)			
$\leq 12$ weeks	28 (77.8%)		
> 12 weeks	8 (22.2%)		

#### Table 1: Demographic and Clinical Characteristics of the Included Women. 410

411 Data were expressed as median(range) or frequency %.

\*Chi-square/Fisher Exact tests compare proportion between groups. \*\*Mann-Whitney U Tests compare the median between groups

P value is considered significant when < 0.05.

412 413 414 415

Group I (Cases): women having a history of two or more miscarriages. Group II (Controls): Healthy controls with no history of miscarriage and at least one uncomplicated full-term pregnancy.

416

VTE: venous thromboembolism. 417

#### 419 Table 2: Laboratory Characteristics of the Included Women

Median (range)	Group I (n=195)	Group II (n=90)	P-Value*
WBC	5.9 (2.47-13.60)	5.79 (2.47-10.50)	0.096
HB (g/dl)	12 (8.5-14.6)	12.2 (8.5-13.9)	0.227
Platelets (×10 <sup>3</sup> /µl)	282.70 (161.10-430.20)	290.00 (163.10-430.00)	0.275
Prothrombin time	12.20 (10.1-15.4)	12.20 (10.5-14.5)	0.810
<b>Prothrombin concentration %</b>	103.00 (79.0-174.7)	102.15 (84.0-143.0)	0.794
INR	1.00 (0.83-1.19)	1.00 (0.86-1.15)	0.433
APTT (Sec)	31.60 (22.5-43.1)	31.65 (22.5-43.1)	0.996
TT	18.00 (12.2-35.0)	17.85 (12.2-35.0)	0.290
Fibrinogen	3.20 (1.7-6.2)	3.20 (2.0-5.8)	0.648

Data were expressed as frequency and %. \*Mann Whitney U Test was used to compare the median difference between the two groups.

420 421 422 423 424 P value is considered significant when < 0.05.

Group I (Cases): women having a history of two or more miscarriages. Group II (Controls): Healthy controls with no history of miscarriage and at least one uncomplicated full-term pregnancy.

#### Table 3: Comparison of Prothrombin gene G20210A and MTHFR C677T between Group I and 425 Group II

# 426

Variables	Group I (n=195)	Group II (n=90)	P-value*
Prothrombin gene G20210A			
<ul> <li>Wild G/G</li> </ul>	193 (99.0%)	90 (100.0%)	1.000
<ul> <li>Heteromutant A/G</li> </ul>	2 (1.0%)	0 (0.0%)	
Prothrombin gene G20210A Allele			
• G (Wild)	388 (99.5%)	180 (100.0%)	0.935
<ul> <li>A (Mutant)</li> </ul>	2 (0.5%)	0 (0.0%)	
MTHFR C677T gene			
<ul> <li>Wild C/C</li> </ul>	105 (53.8%)	53 (58.9%)	0.569
<ul> <li>Heteromutant C/T</li> </ul>	65 (33.3%)	29 (32.2%)	
<ul> <li>Homomutant T/T</li> </ul>	25 (12.8%)	8 (8.9%)	
MTHFR C677T gene Allele			
<ul> <li>C (Wild)</li> </ul>	275 (70.5%)	135 (75.0%)	0.269
T (Mutant)	115 (29.5%)	45 (25.0%)	

Data were expressed as frequency and %. \*Chi-square test and Fisher Exact test were used to compare proportion between groups

P value is considered significant when < 0.05.

Group I (Cases): women having a history of two or more miscarriages.

427 428 429 430 431 Group II (Controls): Healthy controls with no history of miscarriage and at least one uncomplicated full-term pregnancy.





434 Fig. (1): Study flow chart.





436 Fig. (2): Comparison of Thrombophilia markers (Protein C, Protein S, and Antithrombin III and

437 Homocysteine) between Group I and Group II.