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Journal of Enzyme Inhibition and Medicinal Chemistry ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/ienz20

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To cite this article: Lamya H. Al-Wahaibi, Mohamed A. Mahmoud, Yaser A. Mostafa, Ali E. Raslan & Bahaa G. M. Youssif (2023) Novel piperine-carboximidamide hybrids: design, synthesis, and antiproliferative activity via a multi-targeted inhibitory pathway, Journal of Enzyme Inhibition and Medicinal Chemistry, 38:1, 376-386, DOI: 10.1080/14756366.2022.2151593

To link to this article: https://doi.org/10.1080/14756366.2022.2151593

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Novel piperine-carboximidamide hybrids: design, synthesis, and

antiproliferative

activity via a multi-targeted inhibitory pathway

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ABSTRACT

A new series of piperine-carboximidamide hybrids VIa-k was developed as a new cytotoxic agent targeting EGFR, BRAF, and CDK2. The antiproliferative effect against four cancer cells was investigated against erlotinib. Hybrids VIc, VIf, VIg, VIi, and VIk have the highest antiproliferative activity. Compounds VIc, VIf, VIg, VIi, and VIk inhibited EGFR with IC₅₀ values ranging from 96 to 127 nM. Compounds VIf and VIk had the most potent inhibitory activity as BRAF_{V600E} (IC₅₀ ¼ 49 and 40 nM, respectively) and were discovered to be potent inhibitors of cancer cell proliferation (GI₅₀ ¼ 44 and 35 nM against four cancer cell lines, respectively). Compound VIk, the most effective derivative as an antiproliferative agent, demonstrated potent anti-CDK2 action with an IC₅₀ value of 12 nM, which is 1.5-fold more potent than the reference dinaciclib. Finally, VIc, VIf, and VIk have a high capacity to inhibit LOX-IMVI cell line survival. GRAPHICAL ABSTRACT

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Vlk MTT assay (4 cancer cell lines) A-549; IC50 = 32 nM MCF-7; IC50 = 35 nM HT-29; IC50 = 38 nM Panc-1; IC50 = 36 nM EGFR inhibition: IC50 = 127 nM BRAFV600E inhibition, IC50 = 40 nM CDK2 inhibiton. IC50 = 12 nM LOX-IMVI melanoma IC50 = 1.05 µM **Cell Viability Assay** (89%) HIGHLIGHTS _ A series of piperine-carboximidamide hybrids VIa-k was designed and synthesised. _ The new compounds were evaluated as antiproliferative agent targeting EGFR, BRAF, and CDK2. _ Antiproliferative activities were evaluated against four human cancer cell lines. _ Molecular docking studies were carried out against EGFR, mutated BRAF and CDK2-TK. In silico ADME/pharmacokinetic analysis was considered. ARTICLE HISTORY Received 27 August 2022 Revised 12 November 2022 Accepted 20 November 2022 **KEYWORDS** Piperine; amidoxime; hybridisation: antiproliferative: kinases CONTACT Bahaa G. M. Youssif bahaa.youssif@pharm.aun.edu.eg, bgyoussif@ju.edu.sa Pharmaceutical Organic Chemistry Department, Faculty of Pharmacy, Assiut University, Assiut 71526, Egypt; Lamya H. Al-Wahaibi Ihalwahaibi@pnu.edu.sa Department of Chemistry, College of Sciences, Princess Nourah bint Abdulrahman University, Riyadh, Saudi Arabia _Additional Affiliation: Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Badr University, Assiut, Egypt. Supplemental data for this article can be accessed online at https://doi.org/10.1080/14756366.2022.2151593 2022 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. JOURNAL OF ENZYME INHIBITION AND MEDICINAL CHEMISTRY 2023, VOL. 38, NO. 1, 376-386 https://doi.org/10.1080/14756366.2022.2151593 Introduction Because cancer is one of the world's top causes of death, identifying and developing new effective anticancer drugs is one of the most difficult concerns in drug development₁₋₅. A single-targeted treatment method that results in chemotherapy resistance has recently been extensively described₆. This problem is addressed by a combination treatment that has received clinical approval7. Despite the fact that combination therapy offers the potential for extra and potentially synergistic benefits, it typically results in unanticipated side effects such as increased toxicity. Dual or multi-target medications with lower drug interaction risk, improved pharmacokinetics (PK), and safety profiles might be utilised as an alternative to combination therapy. A dual or multiple target kinases can also help to minimise poor patient adherence, off-target effects, medication interactions, and excessive manufacturing costs8.

Natural products are a major constant source of lead medicines. Natural product-based developments, including anticancer medicines such as docetaxel, topotecan, and etoposide have been

reported_{9,10}. Natural products derived from bacteria, fungi, marine organisms, plants, and animals, as well as natural product-inspired compounds, have shown promising results in clinical studies, including anticancer treatments₁₁. Natural product-derived derivatives are thought to account for more than half of anticancer medicines; around 74% of anticancer drugs are either natural or natural product-inspired compounds₁₂.

Piperine I, Figure 1, is found in black pepper powder fruit and is

widely utilised as a food flavour in a variety of cultures, as well as in many traditional food preservation systems and traditional treatments₁₃. Piperine has been shown to have anticancer effect through a variety of pathways. The substitution of the piperidine moiety with other pharmacophoric groups has been shown to increase activity and potency, as demonstrated by compounds 2 and 39,14. Amidoxime-containing compounds, on the other hand, demonstrated strong pharmacological effects, with anticancer activity topping the list_{15–18}. The amidoxime moiety has been included as carboxylic and ester group bioisosteres, with the goal of developing prodrugs with enhanced pharmacokinetic and pharmacodynamic properties₁₉. Furthermore, amidoxime moiety has demonstrated its capacity to release NO in vivo for the purposes of developing nitric oxide-releasing prodrugs₂₀.

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New hybrids Vla-k
1- Cell Viability assay
2- MTT assaya gainst 4 cancer cell lines
3- Mechanistic studies against 3 kinases
4- LOX-IMVI melanoma cell line assay
Piperidine moiety
Aryl Carboximidamide
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Aryl Carboximidamide
Piperine Backbone
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Piperine Backbone Figure 1. Structures of piperine, compounds 2–5, and newly designed compounds VIa-k. JOURNAL OF ENZYME INHIBITION AND MEDICINAL CHEMISTRY 377 We recently reported on the design, synthesis, and in vitro assessment of a novel series of aryl carboximidamides as dual p38a/BRAFv600E inhibitors. The results revealed that the presence of the carboximidamide moiety is essential for activity, and the best activity correlates with compound 4, Figure 1. Compound 4 showed significant antiproliferative action against a panel of cancer cell lines. It has an IC₅₀ of 0.33 mM and 0.19 mM against BRAFv600E and p38a, respectively₂₁.

Another novel series in which the indazole moiety is substituted at the 6-position with aryl and or/heteroaryl carboximidamide moiety was developed in our lab₂₂. Compound 5 (Figure 1) was the most active synthetic derivative, with a GI50 value of 0.77 mM against the tested four cancer cell lines, compared to the control doxorubicin (GI₅₀ ¼ 1.10 mM). Compound 5 inhibited EGFR, CDK2, and c-Met with IC₅₀ values of 89 nM, 14 nM, and 4.1 nM, respectively. Furthermore, Compound 5 induced apoptosis by increasing cytochrome C levels and activating the intrinsic apoptotic pathway. Motivated by the antiproliferative effects of piperine (I), compounds 2 and 3, as well as the promising antiproliferative action of aryl carboximidamide derivatives 4 and 5, we present here the design, synthesis, and antiproliferative action of a new hybrid scaffold Vla-k (Figure 1), in which the aryl carboximidamide pharmacophore was bound to the piperine backbone using molecular hybridisation approach in order to obtain potent antiproliferative agents. The new hybrids will be tested against four cancer cell lines using the MTT assay to determine their IC50. The most effective hybrids will be tested further for mechanistic action against EGFR. BRAF. and CDK2. Furthermore, the most potent derivatives as BRAF inhibitors will be tested for their inhibitory action against the LOX-IMVI melanoma cell line, which has BRAFV600E kinase overexpression. Finally, docking studies were performed on the most active derivatives against the active sites of EGFR, BRAF, and CDK2. Results and discussion

Chemistry

Scheme 1 depicts the synthetic steps for the key intermediates II, Va-k, and target compounds VIa-k. Piperine I was extracted, purified, and crystallised from natural sources using the methods described₂₃. Piperic acid II was synthesised by hydrolysing piperine I with alcoholic KOH₂₄, as shown in Scheme 1. Amidoximes Va-I were synthesised as described₂₅ and then coupled with piperic acid II using CDI at room temperature for 3 h to obtain the target compounds VIa-k. 1H NMR, 13C NMR, and elemental microanalyses were used to confirm all of the aryl carboximidamides VIa-k. The IR spectrum of VId, a prominent example of this series, shows the presence of peaks at 3498 and 3381 (NH₂), 1760 (C¹/₄O), and 1622 (C¹/₄N). The 1H NMR spectrum of this compound revealed the appearance of new two doublet signals with J¹/₄8.5, 8.7 Hz, corresponding to para disubstituted pattern of amidoxime moiety at d 7.69 and d 7 ppm, respectively, as well as the appearance of new broad singlet NH2 at d 6.76 ppm and finally the appearance of singlet signal of methoxy (OCH₃) group at d 3.8 ppm. The presence of a new carbonyl (C¹/₄O) group signal at 161.4 ppm in the 13C NMR spectra of this molecule confirmed the earlier observations, as did the appearance of a methoxy (OCH₃) group signal at 55.7 ppm.

Biology

In vitro anticancer activity

Cell viability assay. A human mammary gland epithelial (MCF-10A) normal cell line was used to perform the viability test. The viability of compounds VIa-k was determined using the MTT assay after 4 days of incubation with MCF-10A cells_{26,27}. The results o

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b; R = 4-Cl
c; R = 4-Br
d: R = 4-OCH3
e; R = 4-CH3
f; R = 2-CI
q; R = 3-Cl
h; R= 3-Br
i; R = 3,4-di-OCH<sub>3</sub>
j; R = 2,3-phenyl
k: R = 1.3-dioxole
Scheme 1: Synthesis of target compounds VIa-k. Reagents and reaction conditions: i) KOH, Ethanol, reflux 12 h; ii) NH<sub>3</sub>, THF, I<sub>2</sub>, r.t.
3 h, 65-80%; iii) NH2OH, methanol,
NaHCO<sub>3</sub>, reflux 3-5h, 80-90%; iv) CDI, Acetonitrile, r.t. 3 h.
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showed that none of the compounds tested were cytotoxic, and
that majority of the compounds tested at 50 mM had cell viability
more than 85%.
Antiproliferative activity. Compounds VIa-k were investigated for
antiproliferative efficacy against four human cancer cell lines using
the MTT assay and erlotinib as the reference drug: Panc-1 (pancreatic
cancer cell line). MCF-7 (breast cancer cell line). HT-29 (colon
cancer cell line), and A-549 (epithelial cancer cell line)28,29. Table 1
shows the median inhibitory concentration (IC<sub>50</sub>) calculated using
Graph Pad Prism software (Graph Pad Software, San Diego, CA,
USA). The average (GI<sub>50</sub>) versus the four cancer cell lines was
chosen for ease of manipulation.
In general, compounds VIa-k demonstrated strong antiproliferative
activity with GI50 values ranging from 35nM to 100nM when
compared to the reference erlotinib, which had a GI<sub>50</sub> of 33 nM.
The data in Table 1 reveal that VIa-k were more effective against
epithelial cancer cell line (A-549) than breast cancer cell line
(MCF-7).
The 1,3-benzodioxole derivative VIk (R¼1,3-dioxole) was the
most potent of the synthesised derivatives, with a GI50 value of
35 nM, which was comparable to erlotinib and even more potent
than erlotinib in MCF-7 cell line with IC50 1/4 35 nM, while for erlotinib
IC50 1/4 40 nM.
Compound VIi (R1/43,4-di-OMe) ranks second in activity against
the cell lines examined, with a GI50 of 39 nM. The number of the
methoxy groups appears to be important for the antiproliferative
action. Compound VId (R<sup>1</sup>/<sub>4</sub>4-OMe), for example, has a GI<sub>50</sub> of
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54 nM, which is about 1.4-fold less potent than the dimethoxy derivative VIi. Furthermore, compound VIj (R1/42,3-Ph) has a GI50 of 83 nM, which is 2-times lower than VII, indicating that the dimethoxy moiety is the most well tolerated. Compound VIf (R¹/₄2-CI) demonstrated promising antiproliferative activity against the four cancer cell lines, with a GI50 of 44 nM. The antiproliferative action appears to be strongly influenced by the position and/or type of halogen atom. The GI50 of compounds VIb (R¼4-CI) and VIg (R¼3-CI) was 57nM and 47 nM, respectively, indicating that when the chlorine atom is the substituent, position 2 on the phenyl group is the optimum for activity. Furthermore, compound VIc (R¹/₄4-Br) had a GI₅₀ of 50 nM, which is more potent than compound VIb (R¹/₄4-Cl), which had a GI₅₀ of 57 nM, demonstrating that the bromine atom is more tolerated than the chlorine one in terms of antiproliferative action. The unsubstituted derivative, compound VIa (R¼H), was the least potent of all synthesised derivatives, with a GI50 of 100 nM, approximately three times less active as erlotinib, highlighting the relevance of phenyl moiety substitution for antiproliferative action.

EGFR inhibitory activity

 $IC_{50} \pm SEM (nM)$ $VIc 112 \pm 8 57 \pm 6 23 \pm 2$ $VIf 105 \pm 7 49 \pm 5 21 \pm 2$ $VIg 96 \pm 6 63 \pm 6 20 \pm 2$

The EGFR-TK test30 was used to evaluate the inhibitory potency of the most potent derivatives VIc, VIf, VIg, VIi, and VIk against EGFR, and the outcomes are displayed in Table 2. Compounds VIc, VIf, VIg, VIi, and VIk inhibited EGFR with IC50 values ranging from 96 to 127 nM. According to the findings, all the tested derivatives were less potent than erlotinib (IC50 ¼ 80 ± 5 nM). The 3chloro phenyl derivative VIg (R¼3-CI) was the most potent of all synthesised derivatives, with an IC₅₀ value of 96 nM, which was 1.2-fold lower than erlotinib. The results of the EGFR inhibitory assay test indicate that the EGFR may be a potential target for the antiproliferative effect of some compounds. Table 1. Antiproliferative activity of compounds VIa-k and Erlotinib. 0 ŏ õ o N NH₂ Vla-k Comp. Cell viability % R Antiproliferative activity IC50 ± SEM (nM) A-549 MCF-7 Panc-1 HT-29 Average (GI50) Via 89 H 95 ± 10 97 ± 10 104 ± 10 105 ± 10 100 VIb 85 4-Cl 55 ± 6 57 ± 6 59 ± 6 58 ± 6 57 Vic 86 4-Br 45 ± 5 47 ± 5 53 ± 5 55 ± 5 50 Vid 91 4-OCH₃ 50 \pm 5 52 \pm 5 57 \pm 5 57 \pm 5 54 Vie 89 4-CH₃ 69 ± 7 73 ± 7 75 ± 7 75 ± 7 73 VIf 90 2-Cl 41 ± 4 43 ± 4 45 ± 4 47 ± 4 44 Vlg 87 3-Cl 42 ± 5 47 ± 5 49 ± 5 49 ± 5 47 VIh 91 3-Br 61 ± 6 64 ± 6 69 ± 6 67 ± 6 65 Vii 89 3,4-diOMe 36 ± 4 39 ± 3 40 ± 4 40 ± 4 39 VIj 85 2,3-phenyl 78 ± 8 81 ± 8 84 ± 8 88 ± 9 83 Vlk 89 1,3-dioxole 32 ± 3 35 ± 3 36 ± 3 38 ± 3 35 $Erlotinib - - 30 \pm 3 \ 40 \pm 3 \ 30 \pm 3 \ 30 \pm 3 \ 33$ Table 2. Effects of compounds VIc, VIf, VIg, VIi, Vik, Erlotinib, Vemurafenib, and Dinaciclib on EGFR, BRAFV600E, and CDK2. Compound EGFR inhibition IC50 ± SEM (nM) BRAFV600E inhibition IC50 ± SEM (nM) CDK2

Vii $120 \pm 10\ 72 \pm 7\ 17 \pm 2$ Vlk $127 \pm 10\ 40 \pm 4\ 12 \pm 2$ Erlotinib $80 \pm 5\ 60 \pm 5\ ND$ Vemurafenib ND $30 \pm 3\ ND$ Dinaciclib ND ND 20 ± 2 ND: Not Determined JOURNAL OF ENZYME INHIBITION AND MEDICINAL CHEMISTRY 379 BRAF_{V600e} assay

An in vitro study was conducted to evaluate the anti-BRAFv600E of VIc, VIf, VIg, VIi, and VIk₃₁. The enzyme assay revealed that the tested five compounds strongly inhibited BRAFv600E, with IC50 values ranging from 40 to 72nM (See Table 2). In all cases, the IC50 of the investigated compounds are higher than that of vemurafenib (IC50 ¼ 30 ± 3). Compounds VIf and VIk had the most potent inhibitory activity as BRAFv600E (IC50 ¼ 49 and 40 nM, respectively) and were discovered to be potent inhibitors of cancer cell proliferation (GIs0 ¼ 44 and 35 nM, respectively). According to the study's findings, the examined compounds have substantial antiproliferative activity and are effective at inhibiting BRAFv600E.

Cdk2 inhibitory assay

Compounds VIc, VIf, VIg, VIi, and VIk were further investigated for their potential to inhibit the CDK2 enzyme₃₂. The IC₅₀ values are shown in Table 2. According to the results, all compounds inhibited CDK2 with IC₅₀ values ranging from 12nM to 23nM in comparison to the reference dinaciclib (IC₅₀ ¼ 20 nM). Compound VIk, the most effective derivative as an antiproliferative agent, demonstrated potent anti-CDK2 action with an IC₅₀ value of 12 nM, which is 1.5-fold more potent than the reference dinaciclib. On the other hand, compounds VIf, VIg, and VIi exhibited significant activity against CDK2 (IC₅₀ ¼ 21, 20, and 17 nM) being equipotent to dinaciclib. The results of this assay show that the examined compounds have substantial antiproliferative activity and are effective at inhibiting both CDK2 and BRAFv600E.

Lox-IMVI melanoma cell line cytotoxicity assay

The MTT cytotoxicity assay was used to assess the anticancer activity of VIc, VIf, and VIk on the LOX-IMVI melanoma cell line, which comprises BRAF_{V600E} kinase overexpression₃₃. The compounds were evaluated at 5-dose concentrations to ascertain their IC₅₀ values, with staurosporine acting as a control. Table 3 reveals that the derivatives VIc, VIf, and VIk have a high capacity to inhibit LOX-IMVI cell line survival, with IC₅₀ values ranging from 1.05 mM to 1.40 mM. In all cases, the tested compounds were at least 5 times more effective than the reference Staurosporine, which had an IC₅₀ of 7.10 mM. Compounds VIf and VIf demonstrated promising cytotoxic activity against LOX-IMVI melanoma cell line with IC₅₀ values of 1.10 and 1.05 IM, respectively. Molecular docking simulations

To determine the binding affinity and mode of inhibition of the most active derivatives against potential cellular targets of this class of compounds, we ran in-silico molecular docking simulations for compounds VIc, VIf, VIg, VIi, and VIk against EGFR, mutated BRAF, and CDK2 proteins. The results were extremely promising, particularly with compounds VIg, VIi, and VIk against all the cellular targets used in this study.

Starting with the RSCB deposited crystal structure of the EGFR protein having Erlotinib as a co-crystallised ligand (PDB ID: 1M17), simulations revealed good docking score (S¼_5.84 to _6.72 kcal/ mol) with all test compounds as listed in Table S1 (supporting information). Visual inspections of each test molecule's docking poses revealed numerous interactions with various amino acid residues lining the EGFR active site (Figure 2 and Figure S1). Using compound VIg as a model compound (highest docking score; S¼_6.72 kcal/ mol) to examine its docking poses and gain an understanding of the mode of inhibition of this class of compounds within the EGFR Table 3. IC50 cytotoxicity of compounds VIc, VIf, and VIk against LOXIMVI melanoma cell line. Compound LOX-IMVI melanoma IC50 ± SEM (mM) \dot{V} ic 1.40 ± 0.02 Vlf 1.10 ± 0.01 Vlk 1.05 ± 0.01 Staurosporine 7.10 ± 0.05 Figure 2. Schematic representation of docking interactions of compound VIg within EGFR (PDB ID: 1M17) showing two H-bond interactions with PRO 770 and GLY772 as blue-coloured arrows. 380 L. H. AL-WAHAIBI ET AL. protein, we observed H-bond interactions with PRO770 and GLY772 amino acid residues, as shown in Figure 2. Although the docking scores of the test molecules were very promising, their binding interactions with various amino acid residues lining the EGFR active site remained random, and none of them managed to show a binding interaction with key amino acids (GLN767 and MET769) as did co-crystallised ligand. All these findings demonstrated that the EGFR protein is not the best active site to explain the mode of inhibition of such compounds as potential anti-proliferative agents. Docking into BRAF and CDK2 active sites, on the other hand, revealed a significant common bonding interaction between test molecules and the amino acid residues lining these two active sites, as well as at least one or more bonding interactions with key amino acid residues. Additionally, MTT assay results on the LOX-IMVI melanoma cell line, which is known for being enriched with BRAFV600E kinases. showed promising inhibitory results (5-times better than reference compound; staurosporine). As a result of all these intriguing findings regarding CDK2 and BRAFV600E proteins, we chose to investigate the potential mode of interactions within the active sites of these two proteins. As shown in Table S1, compounds VIc, VIf, VIg, VIi, and VIk showed good and comparable docking score and RMSD values to that of co-crystallised ligands. Compound VIk showed the best docking score (S) within its congeners As shown in Figure 3(a), compound VIk managed to have three strong binding interactions in the form of H-bond donor and acceptor bonds, which stabilised its structure within the active site of BRAFV600E kinase protein, which was likely also seen Figure 3. Schematic representation of docking interactions of compound VIk within both BRAFV600E (PDB ID: 4MNF) and CDK2 (PDB ID: 1PYE): (a,b) 2 D Interactions of VIk within 4MNF and 1PYE; respectively; (c) Presumptive binding modes of VIk within 4MNF active site as 3 D diagram: showing three H-bond interactions with Asn 580 and Asp 594 coloured in red-colour. JOURNAL OF ENZYME INHIBITION AND MEDICINAL CHEMISTRY 381 with compound VIf (as shown in Figure S2), confirming how such class of compound could inhibit BRAFV600E protein. On the other hand, visual inspections of the best docking poses obtained with molecular docking of both compounds VIf and VIk within active site of CDK2 protein, revealed their achievement in having H-bonding interactions with two key amino acid residues GLU81 and LEU83 as did the co-crystallised ligand (as shown in Figure 3(b,c) and Figure S3). Additionally, compound VIc showed two H-bonding interactions with another two amino acid residues (ASN 132 and LYS89, as shown in Figure S3) rather than key amino acid residues, and this could explain its lower potency compared to its congers VIf and VIk against CDK2 protein. To summarise, molecular docking simulations revealed a good docking score of such class of compounds within three active sites used in this study, as well as a good idea about the mode of inhibition of these compounds within BRAFV600E and CDK2

active sites.

ADME parameters and drug-likeness computational analysis From our efforts to investigate bioavailability of such class of compounds, we measured physicochemical descriptors, pharmacokinetics, drug-like nature, and medicinal chemistry friendliness of all newly synthesised 11 compounds via SwissADME free web tool₃₄. Starting with role of five (Lipinski's RO5)36,37, all newly synthesised derivatives VIa-k have values for physicochemical within required ranges of candidate drugs, as shown in Table S2 (supporting information). Briefly, having H-bonding centres (either acceptor or donor) helps in enhancing water solubility and Hbond formation with various amino acid residues lining target active sites, in the same way, having acceptable number of rotating bonds helps in adaptation and flexibility alignment of such molecules within target active sites. Additionally, having a partition coefficient (i.e. lipophilicity parameter) within 0.5 to 5, and both TPSA and MR values below 140 and 130; respectively, generally indicates good probability of penetration through cell membranes, gastrointestinal penetration, and hence bioavailability. Fortunately, all the newly synthesised were found to not a substantial substrate for permeability glycoprotein (which is also known as multidrug resistance protein which is responsible for efflux of drugs out of cells and hence reduce their pharmacological efficacy) and having Abbott oral bioavailability score above zero, indicating their high probability of medicinal impact and biological activities in clinical trials₃₈.

Finally, all the compounds pass the five medicinal chemistry filters (Lipinski, Ghose, Veber, Egan, and Muegge) and Pan-assay of interference compounds (PAINS), indicating their specificity for target proteins affected by them and being a drug-candidate for drug discovery and development, as shown in Table S2. Conclusion

In the search for multi-targeted antiproliferative agents, a novel series of piperine-carboximidamide hybrids was designed and synthesised as EGFR, BRAFv600E, and CDK2 inhibitors. The new hybrids demonstrated promising antiproliferative properties. Some new hybrids exhibited strong EGFR and/or BRAFv600E inhibitory activity. Furthermore, some potent antiproliferative derivatives outperformed the reference drug against CDK2. Docking simulations within the active sites of BRAFv600E and CDK2 provided insight into the potential mode of inhibition of these compounds. Furthermore, the new class of compounds follows Lipinski's rule of five, as well as passing all medicinal chemistry filters and having an acceptable bioavailability profile. Following optimisation, these novel hybrids could be considered as potential anticancer agents.

Materials and methods Chemistry

General Details: See Appendix A Plant material

Black Pepper seeds were purchased from the local market and grounded to a powder then kept in dry container. They were ground to 40 mesh and stored in airtight container until further use.

Extraction, isolation and purification of piperine alkaloid from black Pepper

Extraction. The sample of black pepper (100 g) was refluxed with 300 ml of dichloromethane for 2 h. in a round bottom flask₂₃. Attach a water condenser to the top of the flask and allow water to run through it to condense the CH₂Cl₂ vapours. After cooling the flask, use vacuum filtration with a B€uchner funnel and filter paper to filter out the pepper grounds. Wash the grounds with

50 ml CH₂Cl₂. The filtrate resulting from the extraction solutions were concentrated to dryness using rotary evaporator until a dark brown oil is left.

Thin layer chromatography. Thin layer chromatography experiment were carried out on commercial silica coated plates using hexane: ethyl acetate 7:3 (v/v) as a mobile phase (R_1 0.36 for piperine)

Preparative silica gel chromatography. Adsorption of the extract on activated silica gel. Prepared sample of the extract was subjected to a silica gel column (200 g) and eluted with hexane: ethyl acetate system in a gradient manner (increasing polarity) from 15 to 40%. Fractions were collected and monitored by TLC method using hexane: ethyl acetate 7:3 (v/v) as a solvent system and concentrated H₂SO₄ was used as spraying reagent to identify those fractions containing the target compound.

Crystallisation of piperine. The piperine-containing fractions identified by TLC were pooled and the volatiles removed under reduced pressure using rotary evaporator to provide a thick orange oil placed in an ice bath for cooling. The mass of the thus obtained material, being highly enriched in piperine. 50 ml of cold diethyl ether was added with continuous stirring. After stirring for 5 min, the solvent was evaporated again by using rotary evaporator. The oil was placed in an ice bath for it to cool down. 50 ml of cold diethyl ether was added again and left in the refrigerator for about 24 h. After 24 h, the yellowish piperine precipitates were collected and washed with small volumes of cold diethyl ether. Two more re-crystallisation steps were carried out to rise the purity of the piperine.

Compounds II and Va-k were prepared as previously described $24,_{25}$.

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General method for the synthesis of compounds via-k The N,N'-carbonyl diimidazole CDI (2.29 mmol, 2eq.) was added to a stirred solution of piperic acid (II) (1.14 mmol, 1eq.) in dry acetonitrile (10 ml) and stirred for 1 h at room temperature. The appropriate amidoximes Va-I were then added to the resulting mixture and stirred for another 10 h. Following the completion of the reaction (monitored by TLC), The precipitate VIa-k that developed was filtered and then crystallised from acetonitrile.

N'-f[(2E,4E)-5-(2H-1,3-Benzodioxol-5-yl)penta-2,4-dienoyl]oxygbenzene carboximidamide (via). Yield: 0.27 g (70%), yellow solid,

m.p: 175–177 _C, R: 0.26 (Hexane: Ethyl acetate, 2:1, v/v); 1H NMR (400 MHz, d ppm DMSO-d6): 7.73 (d, J¼7.2 Hz, 2H, Ar-H), 7.55–7.44 (m, 5H, Ar-H), 7.29 (s, 1H, Ar-H), 7.04–7.02 (m, 4H, CH¼CH), 6.87 (s, broad, 2H, NH2), 6.06 (s, 2H, O-CH2-O); 13C NMR (100 MHz, d ppm DMSO-d6): 164.35, 156.49, 148.23, 148.01, 144.78, 140.34, 131.79, 130.52, 128.35, 126.76, 124.99, 123.29, 118.96, 108.53, 105.77, 101.41. Anal. Calc. (%) for C19H16N2O4: C, 67.85; H, 4.80; N, 8.33; Found: C, 67.86; H, 4.87; N, 8.46.

N'-f[(2E,4E)-5-(2H-1,3-Benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-4chlorobenzene-1-carboximidamide (VIb). Yield: 0.08 g (35%), yellow solid, m.p: 157–160 _C, Rt. 0.28 (Hexane: Ethyl acetate, 2:1, v/v); 1H NMR (400 MHz, d ppm DMSO-d₆): 8.55 (s, 1H, Ar-H), 7.82 (d, J¼8.6 Hz, 2H, Ar-H), 7.72–7.65 (m, 3H, Ar-H), 7.45 (d, J¼8.5 Hz, 1H, Ar-H), 7.25 (t, J¼7.7 Hz, 2H, CH¼CH), 7.14–7.06 (m, 2H, CH¼CH), 7.1 (s, broad, NH₂), 6.09 (s, 2H, O-CH₂-O); 1₃C NMR (100 MHz, d ppm DMSO-d₆): 162.00, 156.64, 152.26, 149.18, 148.85, 148.13, 143.62, 136.88, 135.88, 130.59, 129.99, 129.34, 128.64, 128.53, 128.12, 127.83, 127.15, 124.51, 124.02, 117.78, 116.64, 108.73, 105.90, 101.60. Anal. Calc. (%) for C19H15CIN₂O4: C, 61.55; H, 4.08; N, 7.56. Found: C, 61.86; H, 4.20; N, 7.77. N'-f[(2E,4E)-5-(2H-1,3-Benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-4bromobenzene-1-carboximidamide (VIc). Yield: 0.12 g (42.1%), yellow solid, m.p: 159-161 _C, Rr. 0.29 (Hexane: Ethyl acetate, 2:1, v/v); IR (FTIR): mmax (cm_1) 3491,3358 (NH2), 3134, 2919 (CH), 1769 (C¼O), 1624, 1590 (C¼N and C¼C), 834 (Ar-CH bending); 1H NMR (400 MHz, d ppm DMSO-d₆): 8.55 (s, 1H, Ar-H), 7.82 (s, 1H, Ar-H), 7.82–7.74 (m, 1H, Ar-H), 7.26 (t, J¹/₄7.27 Hz, 2H, CH¹/₄CH), 7.14-7.08 (m, 4H, Ar-H), 7.00 (s, 2H, CH¼CH), 6.97 (s, broad, 2H, NH₂), 6.09 (s, 2H, O-CH₂-O). 13C NMR (100 MHz, d ppm DMSO-d₆): 161.99, 156.67, 152.27, 149.13, 148.86, 148.14, 143.59, 136.92, 132.15, 131.45, 131.02, 130.37, 130.10, 128.88, 127.96, 127.38, 124.49, 124.11, 117.80, 116.63, 108.72, 105.91, 101.59. Anal. Calc. (%) for C19H15BrN2O4: C, 54.96; H, 3.64; N, 6.75. Found: C, 55.12; H, 3.75: N. 6.87. N'-f[(2E,4E)-5-(2H-1,3-Benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-4methoxybenzene-1-carboximidamide (VId). Yield: 0.24 g (72.7%), vellow solid, m.p: 145-148 C, Rf: 0.29 (Hexane: Ethyl acetate, 2:1, v/v); IR (FTIR): mmax (cm_1) 3498,3381 (NH2), 3134, 2978 (CH), 1760 (C1/4O), 1683, 1588 (C1/4N and C1/4C), 827 (Ar-CH bending); 1H NMR (400 MHz, d ppm DMSO-d₆): 8.55 (s, 1H, Ar-H), 7.82 (s, 1H, Ar-H), 7.77 (d, J¼10.9 Hz, 1H, Ar-H), 7.69 (d, J¼8.5 Hz, 2H, Ar-H), 7.14-7.06 (m, 4H, CH¼CH), 7.00 (d, J¼8.7 Hz, 2H, Ar-H), 6.76 (s, broad, 2H, NH2), 6.09 (s, 2H, O-CH2-O), 3.8 (s, 3H, O-CH3). 13C NMR (100 MHz, d ppm DMSO-d₆): 161.50, 157.61, 152.98, 149.64, 144.08, 128.68, 124.99, 124.43, 123.68, 118.23, 117.20, 114.23, 109.22, 106.35, 102.10, 55.84. Anal. Calc. (%) for C20H18N2O5: C, 65.57; H, 4.95; N, 7.65. Found: C, 65.43; H, 5.10; N, 7.77. N'-f[(2E,4E)-5-(2H-1,3-Benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-4methylbenzene-1-carboximidamide (VIe). Yield: 0.20 g (50%), yellow solid, m.p: 185-188 C, Rf: 0.24 (Hexane: Ethyl acetate, 2:1, v/ v); 1H NMR (400 MHz, d ppm DMSO-d6): 7.63 (d, J¹/₄7.76 Hz, 2H, Ar-H), 7.53–7.46 (m, 1H, CH¼CH), 7.28 (s, 1H, Ar-H), 7.25 (d, J¹/₄7.76 Hz, 2H, Ar-H), 7.00 (d, J¹/₄4.36 Hz, 2H, CH¹/₄CH), 7.00 (d, J1/4.36 Hz, 1H, Ar-H), 6.93 (d, J1/47.96 Hz, 1H, Ar-H), 6.81 (s, broad, 2H, NH₂), 6.18 (d, J¹/₄15.4 Hz, 1H, CH¹/₄CH), 6.00 (s, 2H, OCH₂-O), 2.50 (s, 3H, Ar-CH₃). 13C NMR (100 MHz, d ppm DMSO-d₆): 164.40, 156.35, 148.22, 148.06, 144.70, 140.28, 140.15, 130.50, 128.86, 126.65, 125.00, 123.29, 119.01, 108.52, 105.78, 101.40, 20.94. Anal. Calc. (%) for C₂₀H₁₈N₂O₄: C, 68.56; H, 5.18; N, 8.00. Found: C, 68.80; H, 5.30; N, 8.17. N'-f[(2E,4E)-5-(2H-1,3-benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-2chlorobenzene-1-carboximidamide (VIf). Yield: 0.134 g (63%), yellow solid, m.p: 165-167 _C, Rf: 0.26 (Hexane: Ethyl acetate, 2:1, v/v); IR (FTIR): mmax (cm_1) 3491,3358 (NH2), 3134, 2919 (CH), 1769 (C¼O), 1624, 1590 (C¼N and C¼C), 834 (Ar-CH bending); 1H NMR (400 MHz, d ppm DMSO-d6): 8.54 (s, 1H, Ar-H), 7.57–7.47 (m, 4H, Ar-H), 7.43 (d, J¼7.56 Hz, 1H, Ar-H), 7.26 (d, J¼8.2 Hz, 1H, Ar-H), 7.25 (d, J¼15.4 Hz, 1H, CH¼CH) , 7.14-7.07 (m, 3H, CH1/4CH), 6.97 (s, broad, 2H, NH2), 6.08 (s, 2H, O-CH2-O). 13C NMR (100 MHz, d ppm DMSO-d₆): 162.03, 156.68, 152.22, 149.17, 148.84, 148.13, 143.57, 136.92, 132.12, 131.12, 130.50, 129.89, 129.50, 126.97, 124.44, 123.57, 117.99, 116.46, 108.79, 105.87,

101.42. Anal. Calc. (%) for C19H15ClN2O4: C, 61.55; H, 4.08; N, 7.56. Found: C, 61.67; H, 4.23; N, 7.77.

N'-f[(2E,4E)-5-(2H-1,3-benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-3chlorobenzene-1-carboximidamide (VIg). Yield: 0.08 g (44.4%), yellow solid, m.p: 163–165 _C, Rr: 0.26 (Hexane: Ethyl acetate, 2:1, v/v); 1H NMR (400 MHz, d ppm DMSO-d6): 8.28 (s, 1H, Ar-H), 7.65–7.59 (m, 2H, Ar-H), 7.15–7.06 (m,2H, CH¼CH), 7.08 (s, broad, 2H, NH₂), 6.95 (d, J¼7.6 Hz, 1H, Ar-H), 6.08 (s, 2H, O-CH₂-O). 13C NMR (100 MHz, d ppm DMSO-d6):, 162.33, 149.95, 149.35, 144.26, 130.96, 130.66, 124.96, 124.36, 118.28, 117.06, 109.70, 106.28, 101.87, Anal. Calc. (%) for C19H15CIN2O4: C, 61.55; H, 4.08; N, 7.56. Found: C, 61.43; H, 4.26; N, 7.83.

N'-f[(2E,4E)-5-(2H-1,3-benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-3bromobenzene-1-carboximidamide (VIh). Yield: 0.34 g (72.34%), yellow solid, m.p: 141–143 _C, Rr: 0.28 (Hexane: Ethyl acetate, 2:1, v/v); IR (FTIR): m_{max} (cm_1) 3486, 3348 (NH2), 3134, 2916 (CH), 1778 (C¼O), 1622, 1589 (C¼N and C¼C), 988, 830, 761 (Ar-CH bending); 1H NMR (400 MHz, d ppm DMSO-d₆): 8.55 (s, 1H, Ar-H), 7.92 (s, 2H, Ar-H), 7.75 (t, J¼6.74 Hz, 4H, Ar-H), 7.45 (t, J¼7.9 Hz, 1H, Ar-H), 7.45 (t, J¼7.9 Hz, 1H, CH¼CH), 7.27 (t, J¼7.7 Hz, 1H, Ar-H), 7.27 (t, J¼7.7 Hz, 1H, CH¼CH), 7.13–7.07 (m, 2H, CH¼CH), 6.98 (s, broad, 2H, NH2), 6.09 (s, 2H, O-CH₂-O). 13C NMR (100 MHz, d ppm DMSO-d₆): 162.33, 156.64, 152.46, 149.65, 148.36, 143.95, 133.85, 131.27, 130.36, 129.67, 126.25, 124.96, 124.36, 122.08, 118.28, 117.37, 109.17, 106.28, 101.87. Anal. Calc. (%) for C19H15BrN₂O₄: C, 54.96; H, 3.64; N, 6.75. Found: C, 55.06; H, 3.79; N, 6.97.

N'-f[(2E,4E)-5-(2H-1,3-benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-3,4dimethoxybenzene-1-carboximidamide (VIi). Yield: 0.134 g (37.2%), yellow solid, m.p: 147–149 _C, Rf: 0.24 (Hexane: Ethyl acetate, 2:1, v/v); IR (FTIR): m_{max} (cm_1) 3490, 3378 (NH₂), 3079, 2938 (CH), 1757 (C¼O), 1623, 1585 (C¼N and C¼C), 766 (Ar-CH bending); 1H NMR (400 MHz, d ppm DMSO-d₆): 8.56 (s, 1H, Ar-H), 7.36–7.29 (m, JOURNAL OF ENZYME INHIBITION AND MEDICINAL CHEMISTRY 383 4H, Ar-H), 7.25 (d, J¼7.33 Hz, 1H, Ar-H), 7.15–7.08 (m, 2H, CH¼CH), 7.03 (d, J¼8.34 Hz, 2H, CH¼CH), 6.81 (s, broad, 2H, NH2), 6.09 (s, 2H, O-CH₂-O), 3.81 (s, 6H, -(OCH₃)₂). 1₃C NMR (100 MHz, d ppm DMSO-d₆): 162.14, 158.69, 157.38, 152.55, 151.32, 150.63, 149.40, 148.71, 148.10, 125.69, 123.16, 119.63, 117.71, 116.79, 111.95, 111.03, 110.04, 109.12, 55.70. Anal. Calc. (%) for C₂₁H₂₀N₂O₆: C, 63.63; H, 5.09; N, 7.07. Found: C, 63.50; H, 5.29; N, 7.17.

N'-f[(2E,4E)-5-(2H-1,3-benzodioxol-5-yl)penta-2,4-dienoyl]oxyg naphthalene -1-carboximidamide (VIj). Yield: 0.148 g (70%), yellow solid, m.p: 170–172 _C, R: 0.26 (Hexane: Ethyl acetate, 2:1, v/v); 1H NMR (400 MHz, d ppm DMSO-d₆): 8.56 (s, 1H, Ar-H), 8.00 (m, 2H, Ar-H), 7.59 (m, 5H, Ar-H), 7.31 (s, 1H, Ar-H), 7.07 (m, 3H, CH¹/₄CH), 7.05 (s, broad, 2H, NH2), 6.95 (d, J¹/₄8.0 Hz, 1H, Ar-H), 6.22 (d, J¹/₄15.3 Hz, 1H, CH¹/₄CH), 6.08 (s, 2H, O-CH₂-O). 1₃C NMR (100 MHz, d ppm DMSO-d₆): 164.84, 157.55, 148.66, 145.25, 140.76, 133.47, 131.27, 130.05, 128.46, 127.47, 127.17, 126.56, 126.25, 125.57, 123.98, 119.57, 109.02, 106.25, 101.87. Anal. Calc. (%) for C₂₃H₁₈N₂O₄: C, 71.49; H, 4.70; N, 7.25. Found: C, 71.55; H, 4.83; N, 7.47.

N'-f[(2E,4E)-5-(2H-1,3-Benzodioxol-5-yl)penta-2,4-dienoyl]oxyg-2Hnaphtho[1,2-d][1,3]dioxole-8-carboximidamide (VIk). Yield: 0.09 g (70%), yellow solid, m.p: 166–168 _C, Rf: 0.27 (Hexane: Ethyl acetate, 2:1, v/v); 1H NMR (400 MHz, d ppm DMSO-d6): 8.41 (s, 1H, Ar-H), 7.79 (s, 1H, Ar-H), 7.4–7.3 (m, 2H, Ar-H), 7.00 (s, broad, 2H, NH2), 6.95(dd, J¼8.3 Hz, 7.7 Hz, 2H, Ar-H), 6.74–6.82 (m, 2H, CH¼CH), 6.68 (d, J¼8 Hz, 1H, CH¼CH), 6.19 (d, J¼15.4 Hz, 1H, CH¼CH), 6.06 (s, 2H, O-CH₂-O), 5.93 (s, 2H, O-CH₂-O). 1₃C NMR (100 MHz, d ppm DMSO-d6): 164.46, 162.16, 158.71, 156.40, 150.04, 149.12, 147.81, 147.20, 144.59, 141.44, 140.44, 135.00, 130.54, 125.40, 125.01, 123.18, 121.57, 120.95, 119.03, 118.04, 110.36, 108.83, 108.14, 107.83, 105.60, 102.07, 101.58, 100.77 . Anal. Calc. (%) for C₂₄H₁₈N₂O₆: C, 66.97; H, 4.22; N, 6.51. Found: C, 67.17; H, 4.40; N, 6.65.

Biology

Cell viability testing and IC50 determination

MTT assay. The MTT assay was used to assess how the synthetic compounds impacted the viability of mammary epithelial cells (MCF-10A)_{26,27}. Appendix A.

Antiproliferative test. The MTT assay was performed using various cell lines in accordance with previously reported procedures_{28,29} to investigate the antiproliferative potential of VIa-k. Refer to Appendix A.

EGFR inhibitory assay. The EGFR-TK test₃₀ was used to evaluate the inhibitory potency of the most potent derivatives VIc, VIf, VIg, VIi, and VIk against EGFR. See Appendix A.

BRAF kinase assay. An in vitro study₃₁ was conducted to evaluate the anti-BRAF_{V600E} of VIc, VIf, VIg, VIi, and Vik. See Appendix A. CDK2-TK assay. Compounds VIc, VIf, VIg, VIi, and VIk were further investigated for their potential to inhibit the CDK2 enzyme₃₂. See Appendix A.

LOX-IMVI melanoma cell line cytotoxicity assay. The MTT cytotoxicity assay was used to assess the anticancer activity of VIc, VIf, and VIk on the LOX-IMVI melanoma cell line, which comprises BRAFV600E kinase overexpression₃₃. Refer to Appendix A.

In-Silico molecular docking simulations

Molecular Operating Environment (MOEVR 2014.0901) software was used to evaluate and examine possible interactions of test molecules (VIc, VIf, VIg, VIi, and VIk) within crystal structure of 3 target proteins: Epidermal growth factor receptor (EGFR; PDB ID: 1M17), BRAFV600E kinase (PDB ID: 4MNF) and cyclin-dependent kinase (CDK2; PDB ID: 1PYE) obtained from RSCB protein data bank₃₄ and results in the form of docking score (S; kcal/mol), docking accuracy expressed as root-mean square deviation (RMSD; Å) and binding interactions with various amino acid residues lining active site (as listed in Table S1). Preparation of structural formulas of test molecules and structure of target proteins were performed as reported elsewhere₃₅. Validation of prepared protein structures was done via re-docking of co-crystallised ligands with their protein crystal-structure obtained from RSCB protein data bank and their docking score and RMSD values were within acceptable range for running docking simulations within target proteins (as listed in Table S1). Docking simulations were performed as docking protocol reported elsewhere and results were reported in Table S135. Visual inspection of produced docking poses (10 poses/molecule) for binding interactions with various amino acid residues lining active site of both 4MNF and 1PYE crystal structure and were listed in Table S1 and represented as 2 D and 3 D diagrams in Figures 1-2, Figures S1-S3.

ADME parameters and drug-likeness computational analysis

SwissADME free web tool for prediction of physicochemical parameters₃₄, pharmacokinetics, and drug-likeness of small molecules was used to measure such parameters of compounds VIa-VIk and results were listed in Table S2. Also, the medicinal chemistry filters (Lipinski, Ghose, Veber, Egan, and Muegge) were applied on all test compounds and none disobey such filters. PAINS (Pan-Assay of Interference Compounds) were used to examine specificity of such class of compounds as possible anticancer agents and all compounds gave zero score as shown in Table S2. Disclosure statement

The author reported no potential conflicts of interest(s). Funding

This work was funded by Princess Nourah bint Abdulrahman University Researchers Supporting Project Number [PNURSP2022R3], Princess Nourah bint Abdulrahman University, Riyadh, Saudi Arabia.

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